

# DNAfectin<sup>Tm</sup> Plus Transfection Reagent



# DNAfectin<sup>™</sup> Plus Transfection Reagent

Cat. No.	Description	Quantity
G2500	DNAfectin™Plus	1.0 ml



#### Storage **Conditions**

Store at 4°C. Do not freeze



### Required **Materials**

DNA (0.2-16 µg)

- Serum-free, antibiotic-free medium
- Microcentrifuge tubes



## **Timing**

Preparation: 10 minutes Incubation: 20 minutes Total Incubation: 12-16 hours



## Description

**abm**'s DNAfectin<sup>TM</sup> Plus is a nanoparticle-based, nonliposomal formulation that enables the efficient transfection of plasmid DNA and short oligonucleotides into a broad range of cells with minimal cytotoxicity. This simple protocol does not require the removal of serum or culture medium, resulting in less variability and low risk of contamination. DNAfectin™ Plus has been shown to transfect a wide variety of primary, adherent and suspension cell lines with high efficiency.



# Transfection

To achieve the maximum transfection efficiency and low cytotoxicity, optimize the transfection conditions by varying cell density along with DNA and DNAfectin<sup>TM</sup> Plus concentrations. Optmization Optimal results have been observed when cells are 80-90% confluent and DNA(µg): DNAfectin™ Plus (µl) ratios are 1:1 to 1:5.

#### Table 1: Reagent Quantities for Different Culture Vessels

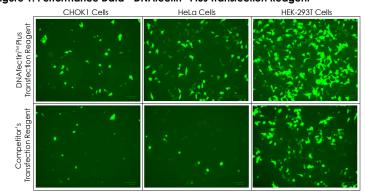
Culture Vessel	Volume of plating medium per well	DNA(µg)	DNAfectin™ Plus (µl)	Transfection medium volume
24-well	500µl	0.2-0.4µg	0.6-1.2µl	50µl
12-well	1ml	0.5-0.8µg	1.5-2.5µl	100µl
6-well	2ml	1.0-2.0µg	3-6µl	200µl
35mm	2ml	1.0-2.0µg	3-6µl	200µl
60mm	5ml	3.0-6.0µg	10-20µl	300µl
10cm	10ml	8.0-16.0µg	25-50µl	500µl

#### Transfection Protocol

Use the following conditions as guidelines to transfect mammalian cells in a 6-well or 35mm dish format. For other culture vessels, please refer to Table 1.

- Plating Cells: 18 to 24 hours prior to transfection, seed the cells at a density such that they are in optimal culture conditions. Incubate the cells at 37°C in a CO<sub>2</sub> incubator until the cells are 70% to 90% confluent at the time of transfection.
- 2. For each transfection sample, prepare the DNAfectin™ Plus-DNA complexes as follows:
  - a) Add 2.0 µg of DNA into 200 µl of serum-free, antibiotic-free medium.
  - b) Warm the DNAfectin<sup>TM</sup> Plus to room temperature and vortex gently before use.
  - c) Add 6.0 µl of the DNAfectin<sup>TM</sup> Plus into the DNA solution from step a). Pipette up and down gently several times to mix the solution
  - d) Incubate for 20 minutes at room temperature to form the DNAfectin™ Plus-DNA complexes. Complexes are stable at room temperature for
- 3. Transfer the DNAfectin<sup>TM</sup> Plus-DNA solution to the cultured cells drop-by-drop to different areas of the culture dish. Gently rock the culture vessel back-andforth and side-to-side to evenly distribute the complexes.
- 4. Incubate for 12-16 hours. It is not necessary to change the culture medium after transfection with DNA fectin™Plus, however, culture medium may be changed between 6-24 hours after transfection for sensitive cell lines.
- 5. Monitor transfection efficiency 24-72 hours post-transfection using relevant assays.

Figure 1: Performance Data - DNAfectin<sup>m</sup> Plus Transfection Reagent



#### Notices and Disclaimers

abm products are intended for laboratory research purposes only, unless noted otherwise. They are not intended for use in humans





